

**HIPPOSPONGIN, A NOVEL FURANOSESTERTERPENE POSSESSING ANTISPASMODIC ACTIVITY
FROM THE OKINAWAN MARINE SPONGE HIPPOSPONGIA SP.**

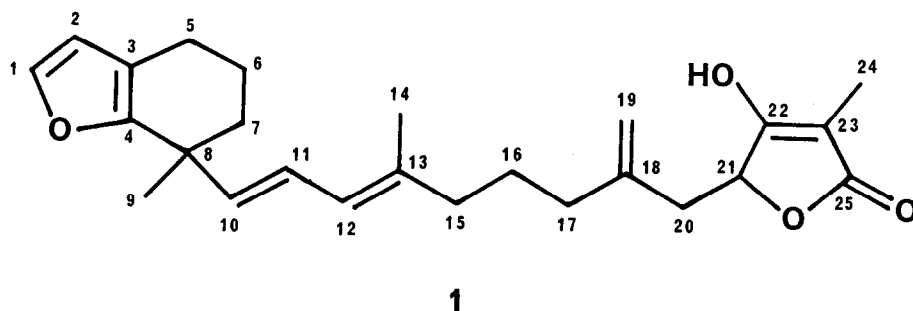
Jun'ichi Kobayashi*, Yasushi Ohizumi, Hideshi Nakamura
Mitsubishi-Kasei Institute of Life Sciences,
11 Minamiooya, Machida, Tokyo 194, Japan
and
Yoshimasa Hirata
Faculty of Pharmacy, Meijo University, Nagoya 468, Japan

Summary: A novel furanosesterterpene, hippospongin, possessing antispasmodic activity has been isolated from the Okinawan marine sponge Hippospongia sp. and its structure determined to be 1 on the basis of the spectral data.

Linear sesterterpenes, characterized by a furan ring at one end and by a tetrone acid at the other, have frequently been encountered in marine sponges^{1,2}. During our studies on bioactive substances in marine invertebrates³⁻⁶, we have isolated from the Okinawan marine sponge Hippospongia sp. a novel furanosesterterpene, named hippospongin (1), which possesses anti-spasmodic activity on the isolated guinea-pig ileum.

The sponge Hippospongia sp. was collected in the Kerama Islands, Okinawa in July 1984. The methanol-toluene (3:1) extract of the sponge was partitioned between toluene and water. The aqueous phase was then extracted with chloroform, ethyl acetate and n-butanol, respectively. The antispasmodic activity was found in the chloroform soluble portion, which was chromatographed on a silica gel column with increasing concentration of methanol in chloroform. Eution with 10% methanol gave hippospongin (1) (0.03% wet weight) as a colorless oil, $[\alpha]_D^{25} +15^\circ$ (c = 5.4, CHCl₃).

The UV $\{\lambda_{\max}^{\text{EtOH}}$ 241 nm (ϵ 25100) $\}$ and IR $\{\nu_{\max}^{\text{KBr}}$ 1740 and 1660 cm⁻¹ $\}$ spectra of 1 were consistent with the presence of a tetrone acid^{7,8}. The molecular formula of C₂₅H₃₂O₄ was established by HREIMS (Δ -1.9 mmu). The 400 MHz ¹H NMR spectrum (Table 1) revealed the α - and β -protons of an α,β -disubstituted furan ring⁹ at δ 7.24 (H-1, d, 1.8 Hz) and 6.16 (H-2, d, 1.8 Hz); three

Table 1. ^1H (400 MHz) and ^{13}C (100 MHz) NMR Data of Hipposponginiol 1.

Position	Proton	m	J (Hz)	Carbon	m
1	7.24 ^a	d	1.8	140.6 ^a	d ^b
2	6.16	d	1.8	110.1	d
3				116.7	s
4				154.4	s
5	2.4	m		22.5	t
6	1.7	m		20.0	t
7	1.7	m		38.4	t
8				38.6	s
9	1.34	s		25.8	q
10	5.60	d	15.2	138.8	d
11	5.95	dd	15.2, 10.6	124.9	d
12	5.77	d	10.6	124.8	d
13				137.0	s
14	1.63	s		16.5	q
15	2.0	m		39.3	t
16	1.5	m		25.8	t
17	2.0	m		35.9	t
18				143.8	s
19	4.89, 4.91	s, s		113.0	t
20	2.26	dd	15.0, 8.2	38.2	t
	2.62	dd	15.0, 3.9		
21	4.79	dd	8.2, 3.9	77.8	d
22				177.5	s
23				97.0	s
24	1.69	s		5.9	q
25				175.6	s

a: δ in ppm in CDCl_3 . b: Multiplicity derived from DEPT data.

protons of conjugated olefins at δ 5.60 (H-10, d, $J=15.2$ Hz), 5.95 (H-11, dd, $J=10.6$ and 15.2 Hz) and 5.77 (H-12, d, $J=10.6$ Hz); terminal vinyl protons at δ 4.89 and 4.91 (H-19, each s); an ABX system at δ 2.26 (H-20, dd, $J=8.2$ and 15.0 Hz), 2.62 (H-20, dd, $J=3.9$ and 15.0 Hz) and 4.79 (H-21, dd, $J=3.9$ and 8.2 Hz); two vinylic methyl groups at δ 1.69 (H-24) and 1.63 (H-14); a sharp three proton singlet at δ 1.34 (H-9), and twelve aliphatic protons at δ 1.5-2.4. The detailed analyses of the COSY data of **1** allowed the assignment of all proton signals, which led to the gross structure **1** for hippospongini. The long-range couplings were observed between H-12 and H-14, H-12 and H-15, H-14 and H-15, H-17 and H-19, H-19 and H-20, and H-21 and H-24, respectively, which supported the presence of partial structures $\text{CH}=\text{C}(\text{CH}_3)-\text{CH}_2$, $\text{CH}_2-\text{C}(\text{=CH}_2)-\text{CH}_2$, and of a tetronic acid moiety. The $\Delta^{10,11}$ double bond was deduced by ^1H NMR data ($J_{10-11}=15.2$ Hz) as E oriented. The E geometry of $\Delta^{12,13}$ double bond was assigned by NOE experiments, in which irradiations of H-14 and H-15 enhanced the signals of H-11 (+12%) and H-12 (+5%), respectively. This was also supported by ^{13}C NMR signals of C-14 (16.5) and C-15 (39.5)¹⁰. Further support of structure assigned was derived from EI mass fragments at m/e 135 ($\text{M}-\text{C}_9\text{H}_{11}\text{O}$)⁺ and 201 ($\text{M}-\text{C}_{14}\text{H}_{17}\text{O}$). The ^{13}C chemical shifts of the tetronic acid moiety (C-21, δ 77.8; C-22, 177.5; C-23, 97.0; C-24, 5.9; C-25, 175.6) of **1** agreed well with those of palinulin⁸, a linear sesterterpene tetronic acid. The cyclohexenofuran ring was assigned from ^{13}C chemical shifts reported for cyclic furanoterpenes^{9,11,12} having ring systems similar to that of **1**. This assignment was supported by NOE experiments, in which irradiations of H-5 enhanced both signals of H-2 (+1%) and H-6 (+3%), while irradiations of H-7 and H-9 caused NOE enhancement of H-10 (+5%) and H-11 (+5%), respectively. The assignments of all ^{13}C signals attached to protons were established by the C-H chemical shift correlation experiments. The stereochemistry at C-8 and C-21 remains to be assigned.

Hippospongini is the first sesterterpene containing an isolated cyclohexenofuran ring from marine sponges, while this is also the first isolation of sesterterpene tetronic acid from the family Spongiidae¹. Hippospongini exhibits antispasmodic activity. In the guinea-pig ileum¹³, the contractile responses to carbachol (10^{-7} M) and histamine (10^{-7} M) were abolished by **1** (5×10^{-6} M). Only agelasidines^{5,14}, diterpene and sesquiterpene derivatives containing guanidine and sulfone units have been known as antispasmodic compound from marine organisms. In addition, hippospongini inhibits growth of a Gram-positive bacterium Bacillus subtilis (21-mm zones of inhibition at 100 $\mu\text{g}/\text{disc}$) but not for a Gram-negative bacterium Escherichia coli and a yeast Saccharomyces cerevisiae. The detailed studies on stereochemistry and pharmacological properties of hippospongini are in progress.

Acknowledgements: We thank Dr. T. Hoshino (Mukaishima Marine Biological

Station, Hiroshima University) for his kind identification of the marine sponge, Dr. A. Y. Nosaka (Japan Bruker) and Prof. T. Miyazawa and Mr. K. Wakamatsu (Department of Biophysics and Biochemistry, Faculty of Science, University of Tokyo) for NMR measurements, Mr. Z. Nagahama for his assistance in collecting the marine sponge, and Miss M. Hamashima of this institute for her technical assistance.

References

1. P. R. Bergquist and R. J. Wells: in "Marine Natural Products", Vol. V, p.1, Academic Press, New York, 1983.
2. D. J. Faulkner, Nat. Prod. Rep., **1**, 551 (1984).
3. J. Kobayashi, Y. Ohizumi, H. Nakamura, T. Yamakado, T. Matsuzaki and Y. Hirata, Experientia, **39**, 67 (1983).
4. Y. Ohizumi, A. Kajiwara, H. Nakamura and J. Kobayashi, J. Pharm. Pharmacol., **36**, 785 (1984).
5. H. Nakamura, H. Wu, J. Kobayashi, M. Kobayashi, Y. Ohizumi and Y. Hirata, J. Org. Chem., **50**, 2494 (1985).
6. Y. Nakamura, J. Kobayashi, J. Gilmore, M. Mascial, K. L. Rinehart, Jr., H. Nakamura and Y. Ohizumi, J. Biol. Chem., in press.
7. W. Hofheinz and P. Schönholzer, Helv. Chim. Acta, **60**, 1367 (1977).
8. G. Alfano, G. Cimino and S. De Stefano, Experientia, **35**, 1136 (1979).
9. T. Matsumoto and S. Usui, Chem. Lett., **1978** 105.
10. J. M. Clough and G. Pattenden, J. Chem. Soc. Perkin I, 3011 (1983).
11. R. Kazlauskas, P. T. Murphy, R. J. Wells, J. J. Daly and P. Schönholzer, Tetrahedron Lett., 4951 (1978).
12. G. Cimino, F. Cafieri, L. De Napoli and E. Fattorusso, Tetrahedron Lett., 2041 (1978)
13. J. Kobayashi, H. Nakamura, Y. Hirata and Y. Ohizumi, Toxicon, **20**, 823 (1982).
14. H. Nakamura, H. Wu, J. Kobayashi, Y. Ohizumi, Y. Hirata, T. Higashijima and T. Miyazawa, Tetrahedron Lett., **24**, 4105 (1983).

(Received in Japan 4 March 1986)